

Challenges and Solutions in the Treatment of CRE infection in Taiwan

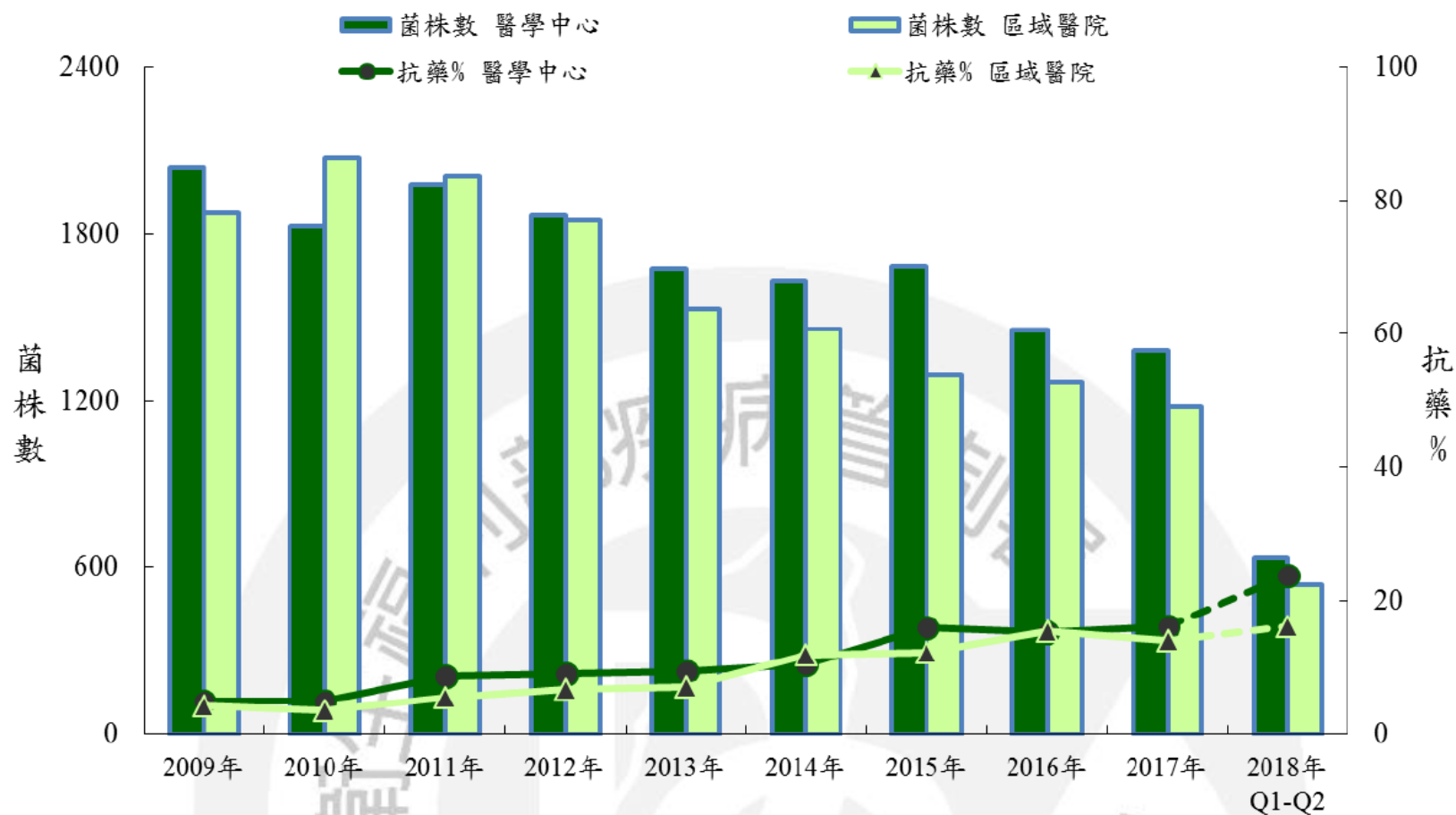
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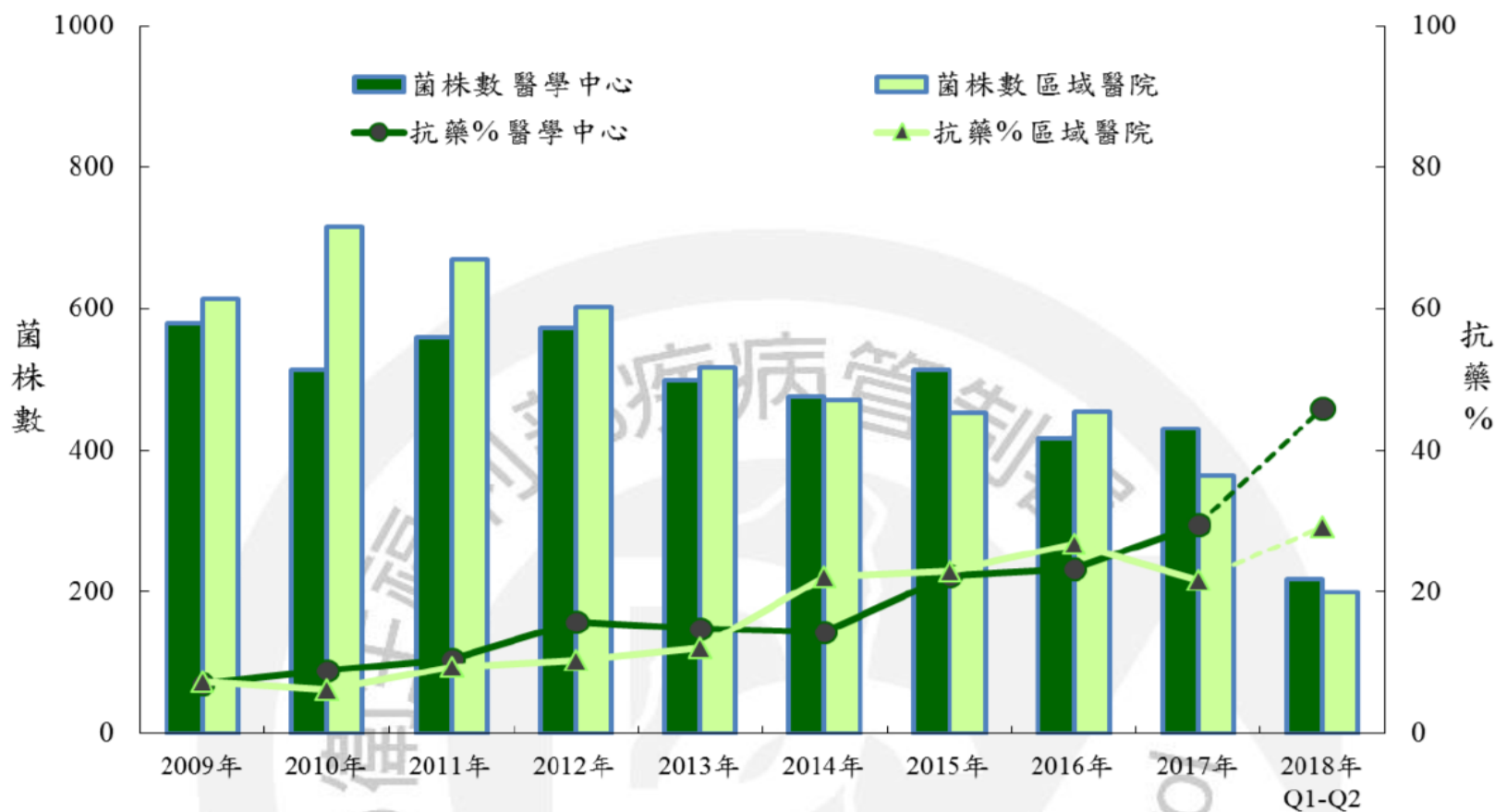
12:10 – 12:40 8 Dec., 2019

CRE = Carbapenem-Resistant Enterobacteriaceae

2009 至 2018 年第 2 季醫學中心及區域醫院加護病房醫療照護相關感染個案分離之 *Enterobacteriaceae* 對 carbapenem 類抗生素具抗藥性(CRE)百分比分布如圖 7。醫學中心加護病房 CRE 比率由 2009 年的 5.0% 增加至 2018 年第 2 季 23.7%；區域醫院則由 4.2% 上升至 16.0%。



2009 至 2018 年第 2 季醫學中心及區域醫院加護病房醫療照護相關感染個案分離之 *Klebsiella pneumoniae* 對 carbapenem 類抗生素具抗藥性(CRKP)百分比分布如圖 11。醫學中心加護病房之 CRKP 比率由 2009 年的 7.1%增加至 2018 年第 2 季 45.9%；區域醫院則由 7.3%上升至 29.1%。



Impact of CRE

- **US in 2013:** 4% of acute hospital, 18% of long-term acute-care facility
- **Mortality** rate of 40-50% in blood-stream infection of CRE-K.P.
 - Underlying severity
 - Delayed administration of appropriate antimicrobial therapy
- **Factors:** elderly, immunosuppressed, chronically, critically ill, health-care exposure, primary immunodeficiency, solid tumors, lymphoma, organ and bone marrow transplant

MMWR Morb Mortal Wkly Rep 2013;62:165. Patel G. Infect Control Hosp epidemiol 2008;29:1099. Cleve Clin J Med 2013;80:225. Sci Transl Med 2012;4:148. Viale P Expert Rev Anti Infect Ther 2013;11:1053. Zarkotou O. Clin Microbiol Infect 2011;17:1798

Mechanisms of Carbapenem Resistance in Enterobacteriaceae

- Production of extended-spectrum β -lactamase (ESBL) and/or AmpC β -lactamase +/- loss of outer membrane protein or upregulation of **efflux pump**: CR- *E. coli*, CR- *Enterobacter* spp. in Taiwan
- Production of **carbapenemases** : CR- *K. pneumoniae* in Taiwan

Pfeifer *Int J Med Microbiol* 2010;300:371. Bush K *Curr Opin Microbiol* 2010;13:558. *Future Microbiol* 2007;2:501. Ma L *BMC Infect Dis* 2013;13:599

Mechanisms of Drug Resistance in Enterobacteriaceae

- Tigecycline-resistant *K. pneumoniae*: ramR deficiency and/or widespread mutated tet (A)
- Fosfomycin-resistant genes: FosA subtypes and fosC2
- Colistin-resistant genes: mobilized colistin resistance (*mcr-1*)

Chiu SK. Antimicrob Agents Chemother 2017;61:e00391. Yang TY, J Microbiol Immunol Infect 2017;10:006

Amber classification

A, B, C, D

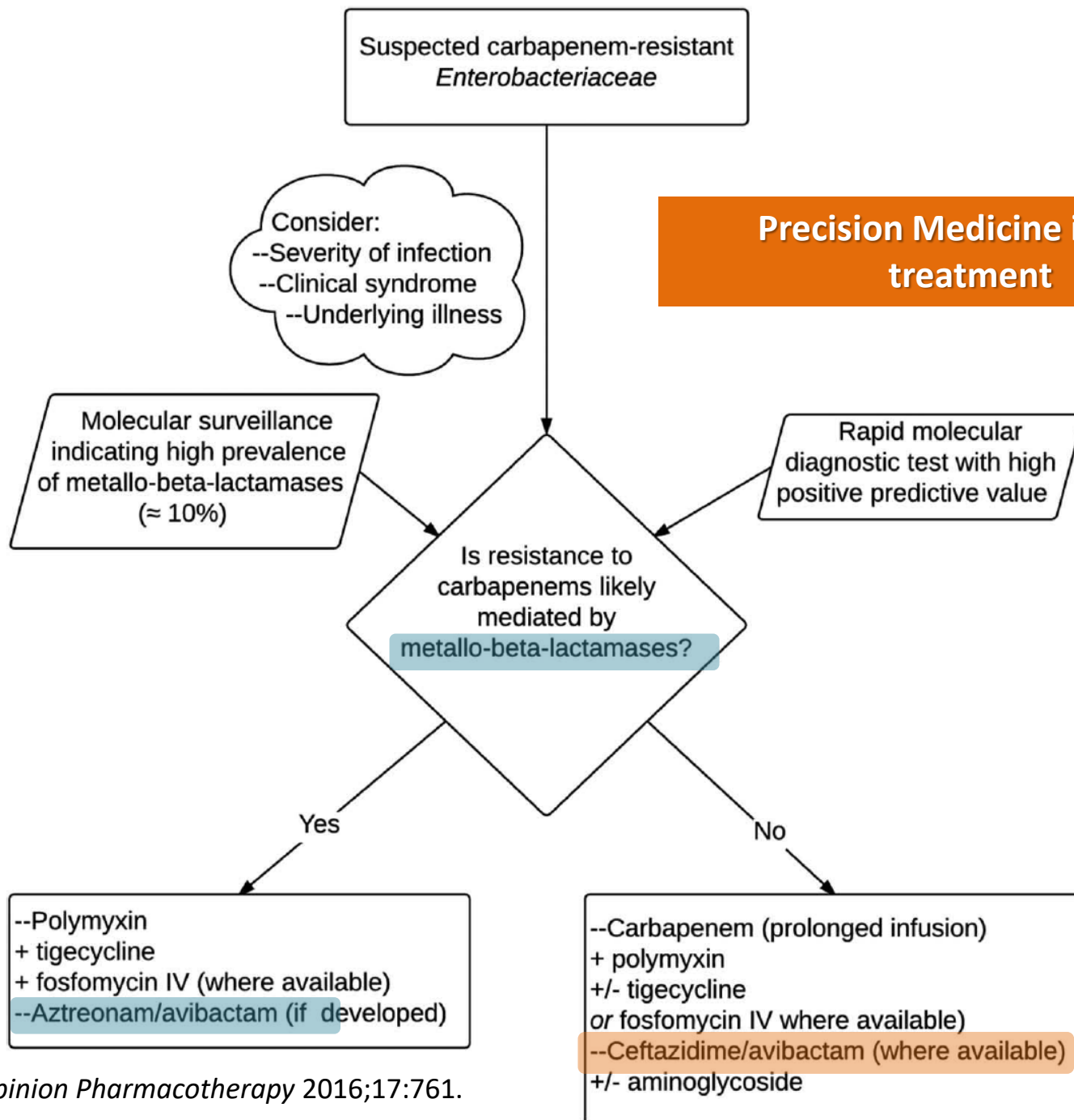
Enzyme/ambler classification	Common genetic platform	Species distribution in <i>Enterobacteriaceae</i>
KPC (<i>Klebsiella pneumoniae</i> carbapenemase)/class A	<i>K. pneumoniae</i> sequence type 258 and ST11 inter alia, transposon Tn4401x	<i>Klebsiella pneumoniae</i> , <i>Escherichia coli</i> , <i>Enterobacter</i> sp.; diverse <i>Enterobacteriaceae</i>
NDM (New Delhi metallo-beta-lactamase)/class B	Various plasmid types	<i>K. pneumoniae</i> and <i>E. coli</i> predominantly; diverse <i>Enterobacteriaceae</i>
OXA-48 (oxacillinase)/class D	Incl/M-type plasmid	<i>K. pneumoniae</i> predominantly, diverse <i>Enterobacteriaceae</i>
VIM (Verona integron-encoded metallo-beta-lactamase)/class B IMP/class B	Gene cassettes in class 1 integrons	<i>K. pneumoniae</i>
SME/class A DHA-1/class C, in combination with OmpK35/36 loss ACT/class C, in combination with OmpK35/36 loss	Chromosome Plasmid	<i>Serratia marcescens</i> <i>K. pneumoniae</i>
CTX-M-15/class A, in combination with OmpK35/36 loss	Plasmid	<i>K. pneumoniae</i>
SHV-5/class A, in combination with OmpK35/36 loss	Plasmid	<i>K. pneumoniae</i>
GES-5/class A, in combination with OmpK35/36 loss	Self-conjugative plasmid	<i>K. pneumoniae</i>

*Expert opinion on
Pharmacotherapy
2016;17:761*

Major Carbapenemases in *Enterobacteriaceae*

Carbapenemase	KPC	MBLs (NDM, VIM, IMP)	OXA-48
Ambler molecular class	A	B	D
Substrates of hydrolysis	All β -lactams	All β -lactams except for aztreonam	Penicillins and carbapenems
Inhibited by classic β -lactamase inhibitors	Minimally	No	No
Inhibited by avibactam	Yes	No	Yes
Inhibited by vaborbactam	Yes	No	No
Inhibited by relebactam	Yes	No	No
Common species in <i>Enterobacteriaceae</i>	<i>K. pneumoniae</i> , <i>E. coli</i> , <i>Enterobacter</i> spp.	NDM: <i>K. pneumoniae</i> , <i>E. coli</i> VIM: <i>K. pneumoniae</i> IMP: <i>K. pneumoniae</i>	<i>K. pneumoniae</i>

KPC, *Klebsiella pneumoniae* carbapenemase; MBL, metallo- β -lactamase; NDM, New Delhi metallo- β -lactamase; VIM, Verona integrin-encoded metallo- β -lactamase; IMP, imipenemase; OXA, oxacillinase.



RESEARCH ARTICLE

Carbapenem-Nonsusceptible
Enterobacteriaceae in TaiwanJann-Tay Wang¹, Un-In Wu², Tsai-Ling Yang³, Lauderdale³, Mei-Chen Chen³, Shu-Ying L
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OPEN ACCESS

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Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

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Abstract

A total of 1135 carbapenem-resistant (nonsusceptible) *Enterobacteriaceae* (CRE) isolates were recovered between November 2010 and July 2012 (517 from 2010–2011 and 618 from 2012) from 4 hospitals in Taiwan. Carbapenemase-producing *Enterobacteriaceae* (CPE) comprised 5.0% (57 isolates), including 17 KPC-2 (16 *Klebsiella pneumoniae* and 1 *Escherichia coli*), 1 NDM-1 (*K. oxytoca*), 37 IMP-8 (26 *Enterobacter cloacae*, 4 *Citrobacter freundii*, 4 *Raoultella planticola*, 1 *K. pneumoniae*, 1 *E. coli* and 1 *K. oxytoca*), and 2 VIM-1 (1 *E. cloacae*, 1 *E. coli*). The KPC-2-positive *K. pneumoniae* were highly clonal even isolates from different hospitals, and all were ST11. IMP-8 positive *E. cloacae* from the same hospitals showed higher similarity in PFGE pattern than those from different hospitals. A total of 518 CRE isolates (45.6%) were positive for *bla*_{ESBL}, while 704 (62.0%) isolates were positive for *bla*_{AmpC}. Among the 518 CRE isolates, 382 (33.6% overall) of which carried both *bla*_{ESBL} and *bla*_{AmpC}. CTX-M-15 (414, 80.0%) was the most common *bla*_{ESBL}, while DHA (497, 70.6%) and CMY (157, 22.3%) were the most common *bla*_{AmpC}. Co-carriage of *bla*_{ESBL} and *bla*_{AmpC} was detected in 31 (54.4%) and 15 (26.3%) of the 57 CPE, respectively. KPC-2 was the most common carbapenemase detected in *K. pneumoniae* (2.8%), while IMP-8 was the most common in *E. cloacae* (9.7%). All KPC-2-positive CRE were resistant to all three tested carbapenems. However, fourteen of the 37 IMP-8-positive CRE were susceptible to both imipenem and meropenem in vitro. Intra- and inter-hospital spread of KPC-2-producing *K. pneumoniae* and IMP-8-producing *E. cloacae* likely occurred. Although the prevalence of CPE is still low, careful monitoring is urgently needed. Non-susceptibility to ertapenem might need to be considered as one criterion of definition for CRE in areas where IMP type carbapenemase is prevalent.

1135 carbapenem-resistant
Enterobacteriaceae (2010.11-
2012.7, variable sources)

- Carbapenemase: 5.0% (A: 17 KPC-2, B:1 NDM-1, 37 IMP-8, 2 VIM-1)
- *bla*_{ESBL}: 45.6% (414 CTX-M)
- *bla*_{AmpC}: 62.0% (497 DHA, 157 CMY)



ORIGINAL ARTICLE

Clinical features of patients with carbapenem nonsusceptible *Klebsiella pneumoniae* and *Escherichia coli* in intensive care units: A nationwide multicenter study in Taiwan

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KEYWORDS

Carbapenem;
Escherichia coli;

Background: Patients in intensive care units (ICUs) are especially prone to colonization and infection by carbapenem-resistant *Enterobacteriaceae*. We conducted a multicenter investigation to study the clinical and microbiological characteristics of patients with

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66 CR-E (46 infections, 20 colonization, 60 KP, 6 E. coli) (2012, ICU)

- Carbapenemase: 28.8%, 19/66 (A: 14 KPC-2, B:1 NDM-1, 1 IMP-8, 3 VIM-1)
- bal_{AmpC} or ESBL = 71.2%, 47/66

- In-hospital mortality for CRE-infection = 50% (23/46)
- Independent factor of septic shock for 30-day mortality

SCIENTIFIC REPORTS

OPEN

High minimum inhibitory concentration of imipenem as a predictor of fatal outcome in patients with carbapenem non-susceptible *Klebsiella pneumoniae*

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Carbapenem resistance in *Klebsiella pneumoniae* is important because of its increasing prevalence and limited therapeutic options. To investigate the clinical and microbiological characteristics of patients infected or colonized with carbapenem non-susceptible *K. pneumoniae* (CnsKP) in Taiwan, we conducted a retrospective study at Taipei Veterans General Hospital from January 2012 to November 2013. Carbapenem non-susceptibility was defined as a minimum inhibitory concentration (MIC) of ≥ 2 mg/L for imipenem or meropenem. A total of 105 cases with CnsKP were identified: 49 patients with infection and 56 patients with colonization. Thirty-one isolates had genes that encoded carbapenemases (29.5%), including *K. pneumoniae* carbapenemase (KPC)-2 (n = 27), KPC-3 (n = 1), VIM-1 (n = 1) and IMP-8 (n = 2). The in-hospital mortality among patients with CnsKP was 43.8%. A MIC for imipenem ≥ 16 μ g/mL, nasogastric intubation and Acute Physiology and Chronic Health Evaluation II score were independent risk factors for in-hospital mortality for all patients with CnsKP. A MIC for imipenem ≥ 16 μ g/mL was also an independent risk factor for 14-day mortality in patients with CnsKP. In conclusion, a positive culture for CnsKP was associated with high in-hospital mortality. A high imipenem MIC of CnsKP can predispose a patient to a poor prognosis.

Klebsiella pneumoniae is an important causative agent of nosocomial and community-acquired Gram-negative bacteremia. It can cause various infections, including pneumonia, urinary tract infections and intra-abdominal infections^{1,2}. *K. pneumoniae* is also the major pathogenic organism of community-onset pyogenic infection in Taiwan and other Asian countries³⁻⁶. *K. pneumoniae* strains possessing extended spectrum β -lactamases (ESBL) conferring multidrug resistance have been reported worldwide. Carbapenems have been used extensively for severe infections arising from ESBL-producing *Enterobacteriaceae*, thereby imposing selection pressure for carbapenem resistance⁷. One of the major carbapenem resistance mechanisms of *K. pneumoniae* is the acquisition of carbapenemase genes that encode enzymes that are able to hydrolyze carbapenems. Another mechanism is the deficiency of outer-membrane porin expression with an overexpression of β -lactamases that possess very weak affinity for carbapenems⁸.

Over the past decade, the prevalence of carbapenem resistance among *K. pneumoniae* has increased dramatically worldwide⁹. The mortality attributable to carbapenem-resistant *K. pneumoniae* infection varies between 26% and 44%, with the highest mortality reported in patients with bacteremia¹⁰. Studies focusing on

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105 CR-KP (49 infections, 56 colonization) (2012.1 – 2013.12, VGHTPE)

- Carbapenemase: 29.5%, 31/105 (A: 27 KPC-2, 1 KPC-3, B: 2 IMP-8, 1 VIM-1)
- bal_{AmpC} or ESBL = 70.5%, 74/105 (55 Amp-C β -lactamase with all DHA-1, ESBL with 38 CTX-M-9, 12 CTX-M-1, 49 SHV)

- In-hospital mortality for CR-KP infection = 63.3% (31/49)

OPEN

Carbapenem Nonsusceptible *Klebsiella pneumoniae* in Taiwan: Dissemination and Increasing Resistance of Carbapenemase Producers During 2012–2015

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Before 2011, the prevalence rates of carbapenemase-producing *Klebsiella pneumoniae* (CPKP) among carbapenem nonsusceptible *K. pneumoniae* (CnSKP) isolates were below 10% in Taiwan. The study presents the dissemination and increased antimicrobial resistance of CPKP from January 2012 to August 2015, as shown by Taiwanese multicenter surveillance. Isolates with minimum inhibitory concentrations (MICs) of $>1\mu\text{g/mL}$ for imipenem or meropenem were collected, screened for various carbapenemase genes by PCR, and tested for antimicrobial susceptibility. Among 1,457 CnSKP isolates, 1,250 were collected from medical centers. The CnSKP prevalence in medical centers increased by 1.7-fold during the study. Among all CnSKP isolates, 457 were CPKP. The CPKP rate among CnSKP increased by 1.5-fold and reached 36.8% in 2015. The CPKP nonsusceptibility rate to aztreonam, fluoroquinolones, and aminoglycosides increased yearly. Six CPKP isolates carried dual carbapenemase genes. Three Ambler classes were identified in 451 isolates with a single carbapenemase: classes A (315 *bla*_{KPC-2}, 2 *bla*_{KPC-3}, 20 *bla*_{KPC-17}, 2 *bla*_{KPC-34}), B (26 *bla*_{IMP-8}, 2 *bla*_{NDM-1}, 36 *bla*_{VIM-1}), and D (40 *bla*_{OXA-48}). The *bla*_{OXA-48} rate among CPKP increased by 6-fold over three years. Most KPC and OXA-48 producers were ST11. CnSKP was increasingly prevalent, owing to CPKP dissemination. Additionally, CPKP became more resistant during the study period.

Klebsiella pneumoniae is a common cause of bacteremia, pneumonia, urinary tract infection, and liver abscess¹. β -lactam antibiotics are often deemed the primary therapeutic option for these infections². Among the β -lactams, carbapenems are considered the antibiotics of last resort³. Once *K. pneumoniae* isolates become nonsusceptible to carbapenems, they are often resistant to all currently available β -lactams and frequently resistant to non- β -lactam antibiotics⁴. In the clinical context, the emergence of carbapenem non-susceptible *K. pneumoniae* (CnSKP) poses a serious threat to patient survival because CnSKP infections have limited treatment options and are associated with high mortality⁵.

Among the many mechanisms conferring resistance to carbapenems, carbapenemases can efficiently hydrolyze carbapenems and have become an important cause of antimicrobial resistance⁶. Many carbapenemases have

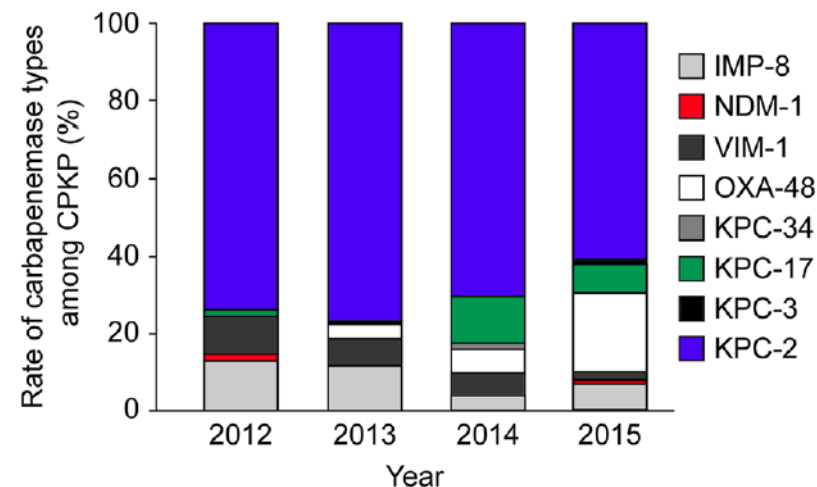
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1457 CR-KP (1250 from medical centers) (2012.1 – 2015.8)

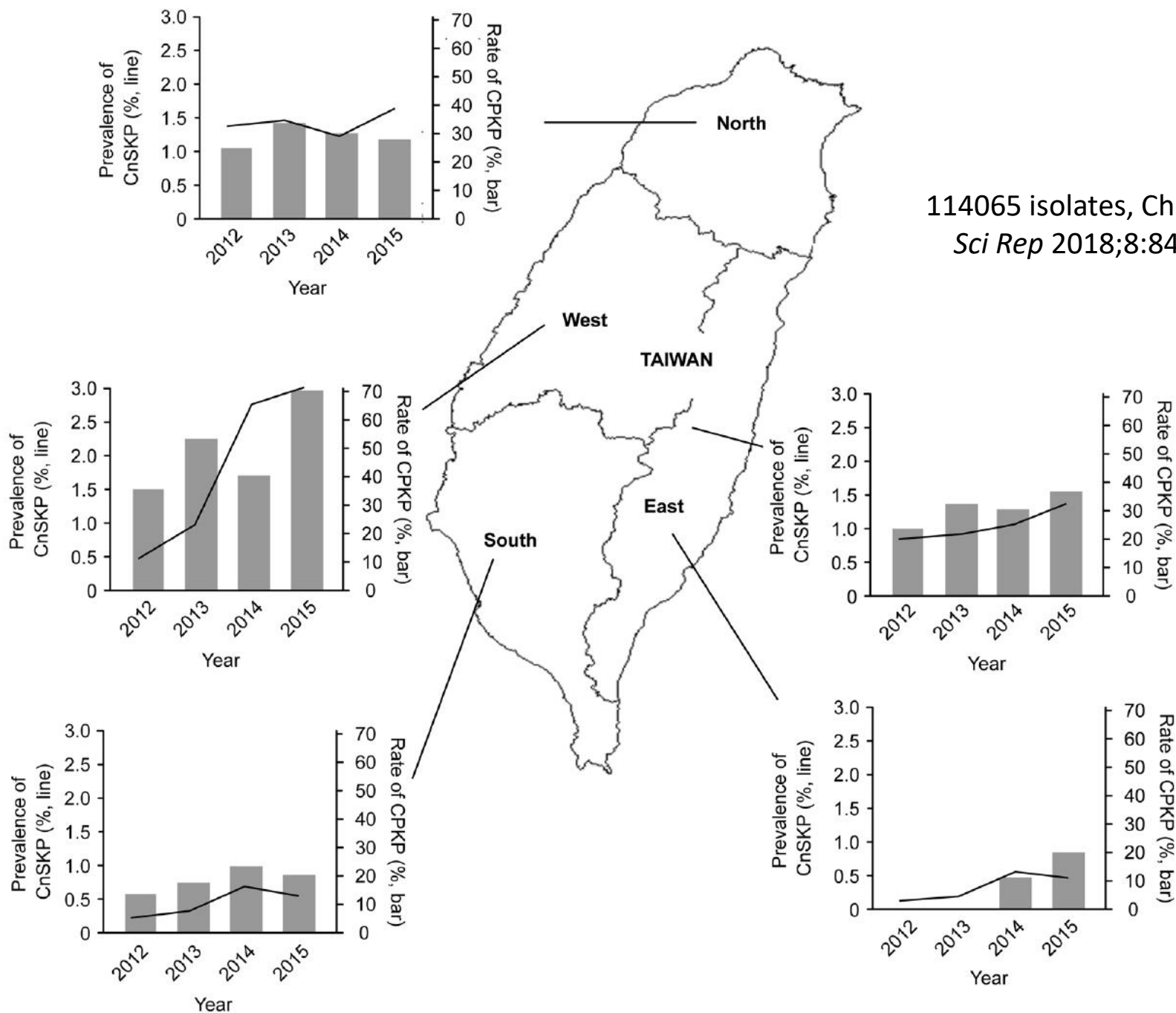
- Carbapenemase: 31.4%, 457/1457 (A: 315 KPC-2, 2 KPC-3, 2 KPC-34;

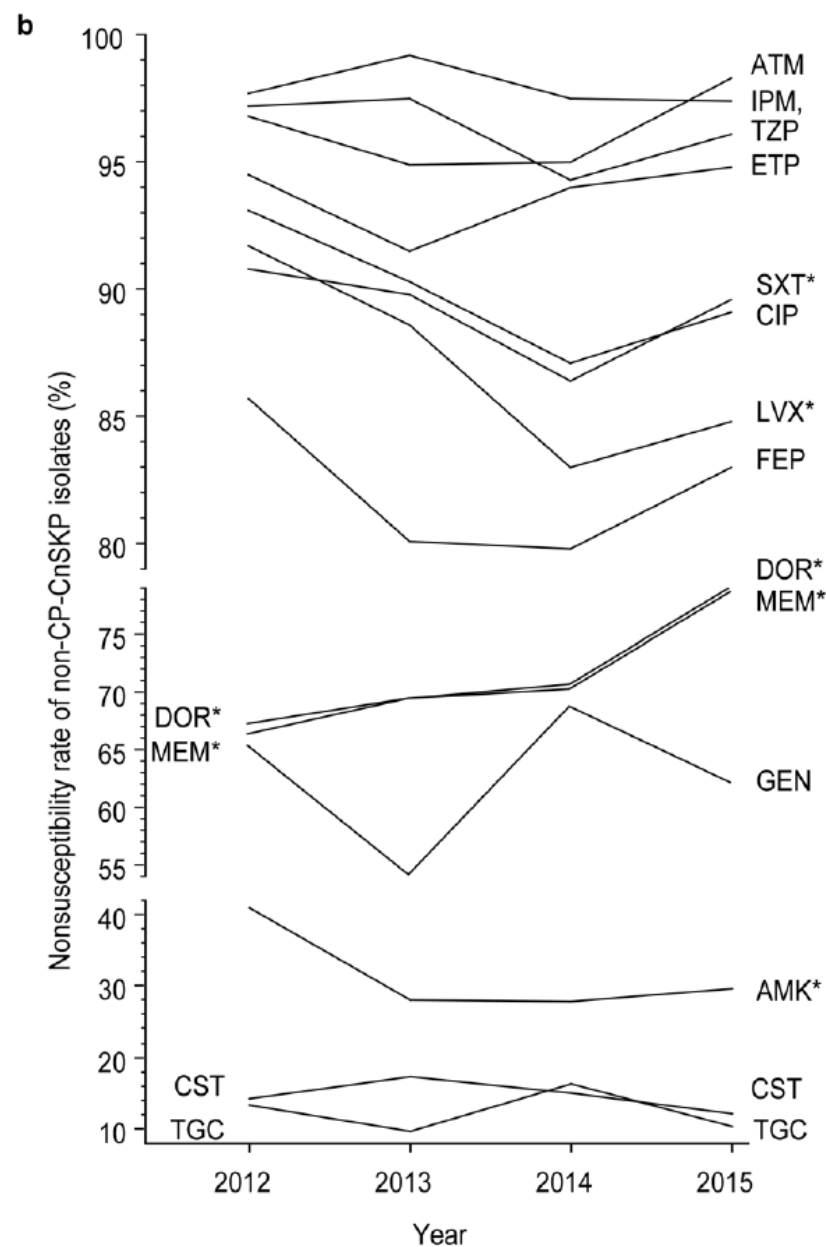
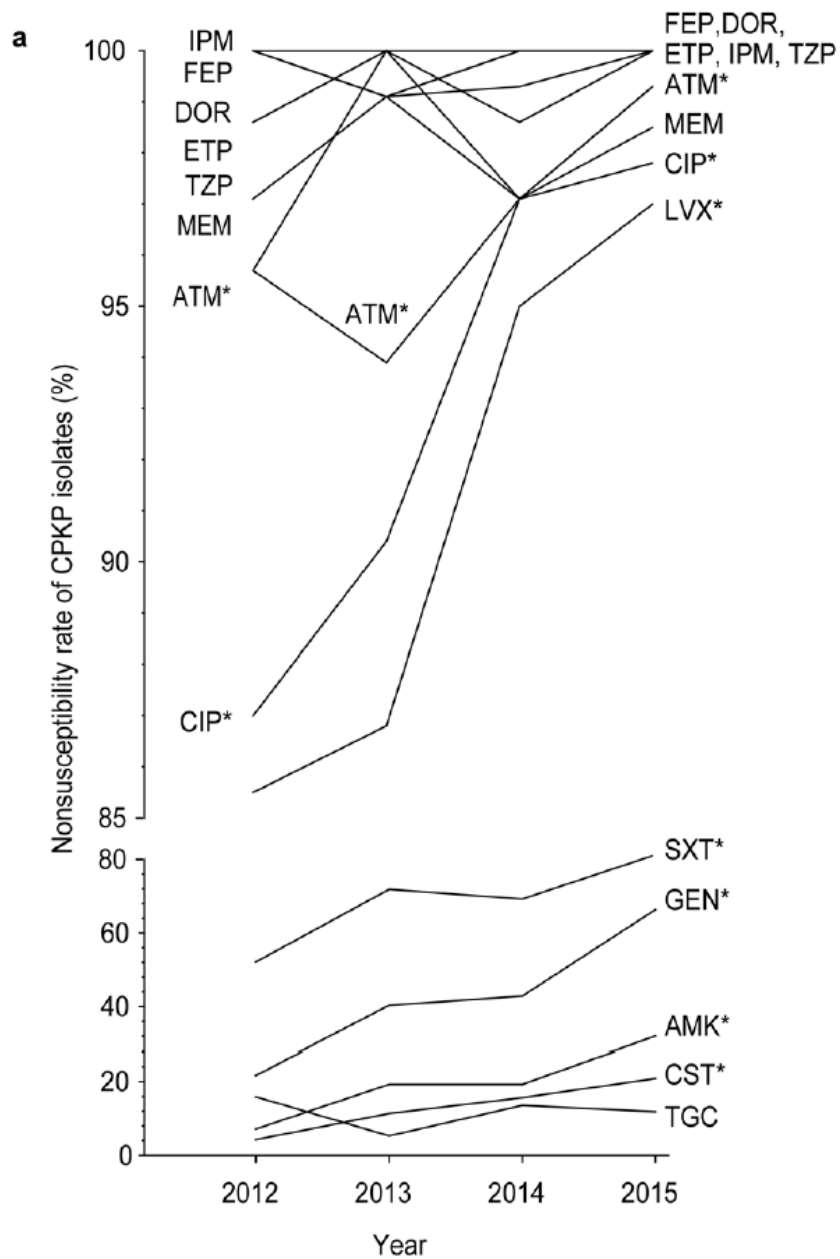
B : 2 NDM-1, 26 IMP-8, 36 VIM-1; D = 40 *bla*_{OXA-48})

- Non-CP-CnSKP = 1000/1457



114065 isolates, Chiu SK.
Sci Rep 2018;8:8468





Resistance mechanisms and molecular epidemiology of carbapenem-nonsusceptible *Escherichia coli* in Taiwan, 2012-2015

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Infection and Drug Resistance

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Purpose: This study aimed to investigate the resistance mechanisms and molecular epidemiology of carbapenem-nonsusceptible *Escherichia coli* (CnsEC) in Taiwan.

Patients and methods: From 2012 to 2015, 237 *E. coli* isolates with minimum inhibitory concentrations of imipenem or meropenem >1 µg/mL were collected in a nationwide surveillance and subjected to polymerase chain reaction (PCR) for carbapenemase, AmpC-type β-lactamase, and extended spectrum β-lactamase (ESBL) genes. We evaluated outer membrane proteins (OmpF and OmpC) loss and conducted multilocus sequence typing and pulsed-field gel electrophoresis (PFGE). Isolates that were resistant to all carbapenems were designated as pan-carbapenem-resistant *E. coli* (pCREC) in this study.

Results: The predominant resistance mechanism of CnsEC in Taiwan was the CMY-2 β-lactamase in combination with OmpF and OmpC loss. Sequence type 131 was the most prevalent type (29.2%). Among 237 CnsEC isolates, 106 (44.7%) isolates were pCREC and 18 (7.59%) produced carbapenemase. The prevalence of carbapenemases increased from 6% in 2012 to 11.36% in 2015. Various carbapenemases including KPC-2, IMP-8, NDM-1, NDM-5, VIM-1, OXA-48, and OXA-181 were identified, with NDM-1 being the most common (38.9%) carbapenemase. Comparison between pCREC and non-pCREC among the non-carbapenemase-producing CnsEC isolates revealed SHV, CMY, co-carriage of SHV and CTX-M and concurrent loss of both OmpF and OmpC were more commonly detected in the pCREC group. PFGE revealed no nationwide clonal spread of carbapenemase-producing *E. coli*.

Conclusion: NDM-1 was the most common carbapenemase and combination of CMY-2 and concurrent OmpF and OmpC porin loss was the most prevalent resistance mechanism in CnsEC in Taiwan.

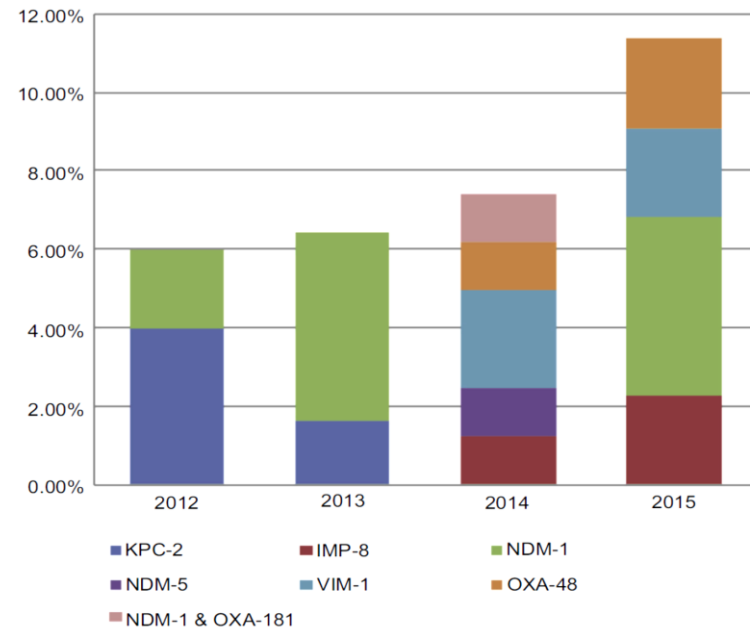
Keywords: multidrug resistance, carbapenemase, Enterobacteriaceae, epidemiology

Introduction

Escherichia coli is one of the most common human pathogen, the major etiology of community-acquired urinary tract infection and a major nosocomial Gram negative bacteria.^{1,2} In the late 1990s, extended spectrum β-lactamase (ESBL)-producing *E. coli* infections emerged.³ Since the worldwide propagation of ESBL-producing *Enterobacteriaceae*, carbapenems have been the prevailing treatment of such infections. Carbapenem resistance in *Enterobacteriaceae* was relatively uncommon before 2000. Nevertheless, the prevalence of carbapenem-resistant *Enterobacteriaceae* (CRE) rose markedly in the following decade.⁴ The United States Centers for Disease Control and Prevention (CDC) listed CRE as an urgent threat that requires intensive monitoring

237 CR-E coli (12 medical centers, 9 LH)
(2012.1 – 2015.9)

- Carbapenemase: 7.59%, 18/237 (A : 3 KPC-2; B : 7 NDM-1, 1 NDM-5, 2 IMP-8, 3 VIM-1; D : 2 bla_{OXA-48})
- Non-CP-CnSKP = 86.5% AmpC, 43.5% ESBL



Susceptibility rates of clinically important bacilli collected from intensive care units against colistin, carbapenems, and other comparative agents: results from Surveillance of Multicenter Antimicrobial Resistance in Taiwan (SMART)

This article was published in the following Dove Medical Press journal:
Infection and Drug Resistance

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Objectives: This study aimed to determine the in vitro susceptibility of common Gram-negative bacilli (GNB) recovered from patients admitted to intensive care units in Taiwan against colistin, carbapenems, and other comparative agents.

Methods: In total, 758 nonduplicate GNB isolates were obtained from clinical ICU patients at seven medical centers in 2016. Minimum inhibitory concentrations were determined using the Vitek 2 susceptibility system. The reference broth microdilution method was performed to determine MICs of colistin. Five main carbapenem-resistant GNB and *mcr-1*–*mcr-5* genes among colistin non-susceptible isolates were determined.

Results: After exclusion 38 *Proteus mirabilis* and 13 *Morganella morganii* isolates, 719 Enterobacteriaceae isolates, 34 (9.4%) isolates were carbapenem-insusceptible, 91 colistin wild type, and three and one *Klebsiella pneumoniae* isolates carried *bla*_{KPC} and *bla*_{OXA48} like, respectively. Carbapenem-insusceptible isolates were found in 23.4% (30 of 128) of isolates of the *Pseudomonas aeruginosa* and *Acinetobacter baumannii* complex isolates, respectively. *mcr-1* was detected in two (1.8%) *Enterobacter cloacae* isolates. Between two methods of susceptibility to colistin were found in 1.5% of *K. pneumoniae*, 4.7% of *P. aeruginosa*, and 10.1% of *A. baumannii* complex isolates.

Conclusion: In this study, 8.7% of Enterobacteriaceae isolates from ICUs were resistant to carbapenem, and *bla*_{KPC} and *bla*_{OXA48} were found among three and one carbapenem-insusceptible *K. pneumoniae* isolates, respectively. Colistin MICs determined by Vitek 2 were not reliable, especially for *E. cloacae* and *A. baumannii* complex isolates.

Keywords: colistin, carbapenems, susceptibility, carbapenemase, *mcr-1*, intensive care units, SMART, *P. aeruginosa*, *A. baumannii*

Introduction

Intensive care units (ICUs) cater to saving the lives of critically ill patients, and their use is rapidly growing worldwide.^{1,2} However, the ICU is also a common place for acquiring nosocomial infections, due to the increasing number of immunocompromised patients and the frequent use of catheters, such as endotracheal tubes, central venous catheters, and Foley catheters.^{3,4} Moreover, the increasing number of multidrug-resistant organisms (MDROs) that cause health-care-acquired infections in the ICU complicates this

SMART 2016, 7 ICU of medical centers

758 GNB (121 *E. coli*, 137 KP, 128 PA, 138 AB...)

- CR-E 9.4%, 34/361; CR-PA 20.3%, CR-AB 62.3%
- Enterobacteriaceae: CR- *E. cloacae* (21.6%), *C. freundii* (20.0%), *K. pneumoniae* (13.1%), *E. coli* (2.5%)
- CP-KP: 3 *bla*_{KPC}, 1 *bla*_{OXA48} (0.5%, 4/758)

In vitro activity of ceftazidime–avibactam, ceftolozane–tazobactam, and other compounds against clinically important Gram-negative bacilli: results from the 2017 Surveillance of Multicenter Antimicrobial Resistance in Taiwan (SMART)

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Objectives: We investigated the in vitro antimicrobial susceptibilities of clinically important Gram-negative bacteria (GNB) from 16 major teaching hospitals in Taiwan in 2017.

Materials and methods: *Escherichia coli* (n=686) and *Klebsiella pneumoniae* bloodstream isolates (n=673), non-typhoid *Salmonella* (NTS; n=221) from fecal samples, and *Neisseria gonorrhoeae* (n=129) collected. Antibiotic minimum inhibitory concentrations (MIC) were determined by microdilution method. Alleles encoding *K. pneumoniae* carbapenemase (KPC), metallo-β-lactamases (NDMs), Verona integrin-encoded metallo-β-lactamases (VIM), and *mcr-1-5* genes were detected by molecular methods.

Results: Five (0.7%) *E. coli* isolates harbored *mcr-1* alleles. Three (0.6%), and one (0.15%) *K. pneumoniae* isolates contained *bla_{NDM}*, respectively. Three (1.4%) NTS and no *Shigella* isolates harbored *mcr-1* alleles. One (10.5%) *K. pneumoniae* isolates displayed non-susceptibility to carbapenems. Phenotypically extended-spectrum β-lactamase (ESBL)-producing isolates showed significantly higher rates of ertapenem, tigecycline, and TAZ NS (40.2%, 16.3%, and 71%–80%, respectively) than non-ESBL-producing isolates (5.4%, 0.7%, and 18%–28%, respectively). All *E. coli* isolates were ceftazidime–avibactam (CAZ–AVB) susceptible. *K. pneumoniae* isolates showed CAZ–AVB NS. Hospital-acquired isolates were significantly less susceptible to ertapenem and CLZ–TAZ than community isolates.

Conclusion: Third-generation cephalosporins remain the mainstay for treating *Shigella*, and gonococcal infections in Taiwan. Hospital-acquired isolates of *K. pneumoniae* are a heavy resistance burden in Taiwan. **Keywords:** Enterobacteriaceae, *Neisseria gonorrhoeae*, carbapenemase, ceftolozane–tazobactam, ceftazidime–avibactam

Introduction

Infections that become septicemic conditions, regardless of the source, are typically associated with

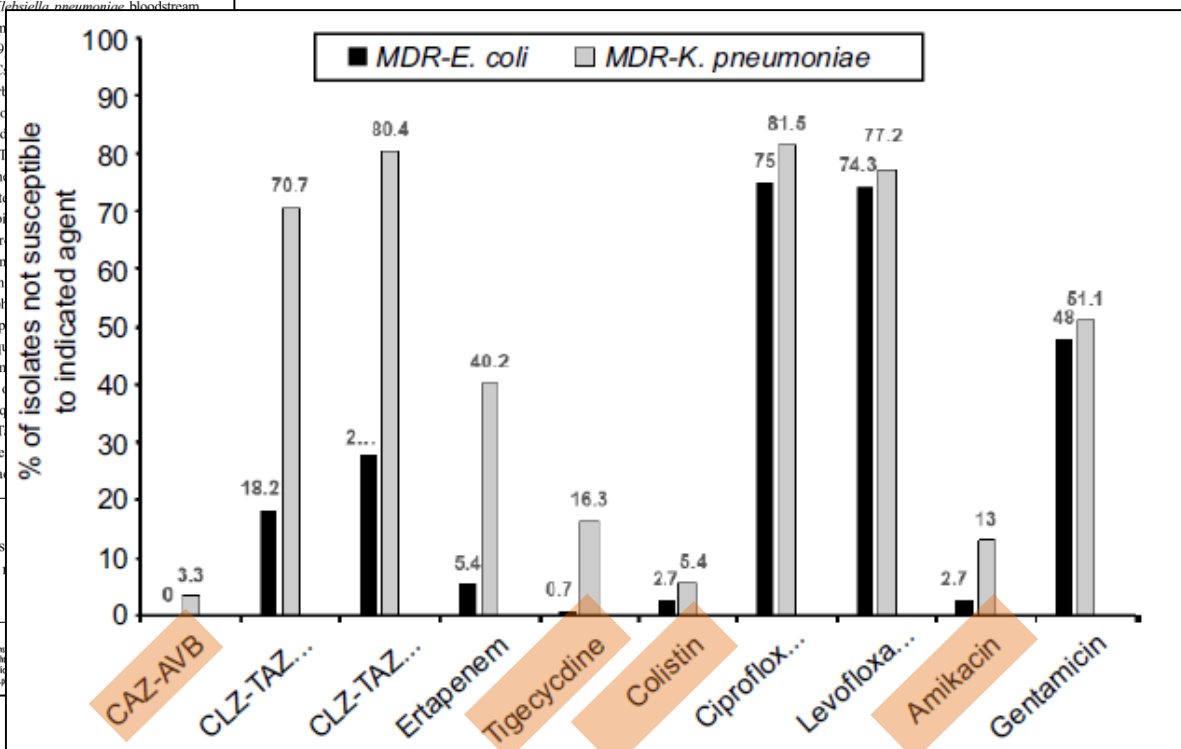
SMART 2017, 16 centers

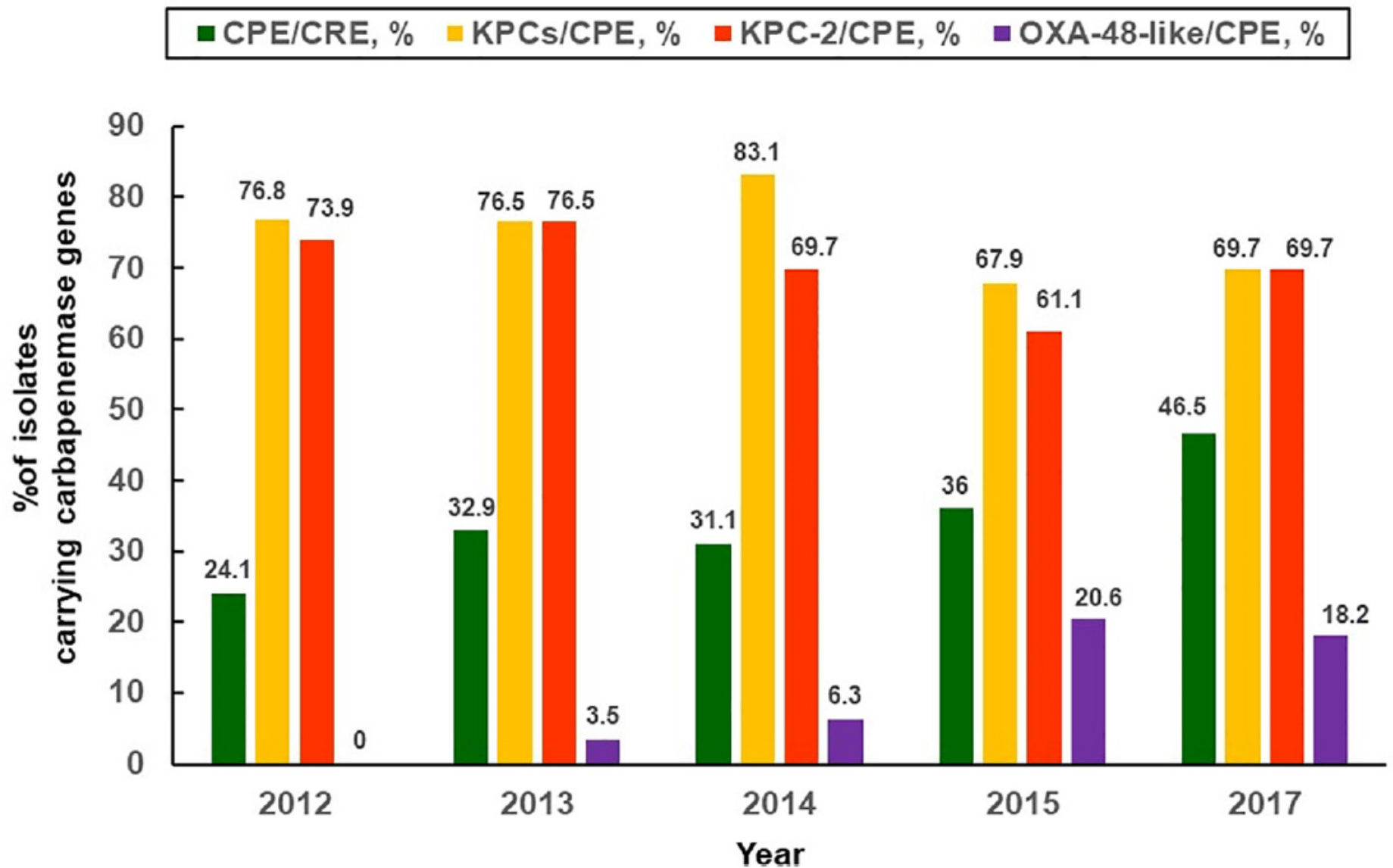
Bloodstream CR-*E. coli* = 2.0%, 14/686

Bloodstream CR-KP = 10.5%, 71/673

85 bloodstream CR-E (14 *E. coli*, 71 KP)

- Carbapenemase: 41.2%, 35/85





Jean SS. *Infect Drug Resist* 2018;11:1983 (SMART **2017**) &
Chiu SK. *Sci Rep* 2018;8:8468 (**2012.1 – 2015.8**) for CR-KP



Nationwide surveillance of antimicrobial resistance among clinically important Gram-negative bacteria, with an emphasis on carbapenem-resistant Enterobacteriaceae and colistin: Results from the Surveillance of Multicenter Antimicrobial Resistance in Taiwan (SMART) in 2018

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2018 SMART, 16 sites

1184 bloodstream GNB (398 *E. coli*, 346 KP, 252 PA, 188 AB, 230 *Salmonella*, 18 *Shigella*)

- CR-*E. coli* 2.8%, CR-KP 9.0%, CR-PA 10.3%, CR-AB 43.6%, CR-*salmonella* spp 0.4%

Prevalence [n (%)]

	<i>E. coli</i> (n = 398)	<i>K. pneumoniae</i> (n = 346)	<i>Salmonella</i> spp. (n = 230)	<i>P. aeruginosa</i> (n = 252)	<i>A. baumannii</i> complex
Carbapenem-non-susceptible	11 (2.8)	31 (9.0)	1 (0.4)	26 (10.3)	82 (43.6)
<i>bla</i> _{KPC}	0	15 (4.3)	0	0	0
<i>bla</i> _{OXA-48}	0	2 (0.6)	0	0	0
<i>bla</i> _{NDM-1}	1 (2.6)	0	0	0	0
<i>bla</i> _{VIM}	0	2 (0.6)	0	0	0
High colistin MIC ^a	2 (0.5)	22 (6.4)	82 (35.7)	22 (8.7)	16 (8.5)
<i>mcr-1</i>	1 (2.6)	2 (0.6)	3 (1.3)	0	0

MIC, minimum inhibitory concentration.

^a An MIC \geq 2 mg/L among Enterobacteriaceae and an MIC \geq 4 mg/L among *P. aeruginosa* and *A. baumannii* complex isolates.

Treatment outcome of non-carbapenemase-producing carbapenem-resistant *Klebsiella pneumoniae* infections: a multi-center study in Taiwan

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Abstract

Carbapenem-resistant *Klebsiella pneumoniae* (CRKP) infections are associated with high mortality, and treatment are usually based on carbapenemase-producing strains. Non-carbapenemase-producing CRKP is of concern but relevant studies are lacking. This nationwide study aimed to evaluate the outcome of antimicrobial therapy in carbapenemase-producing CRKP infections. Patients with non-carbapenemase-producing CRKP infections were treated in hospitals during January 2013 to December 2014 in Taiwan. Carbapenem resistance was defined as reduced sensitivity to imipenem or meropenem. The resistance mechanisms of CRKP were analyzed, and the clinical data of these patients were collected retrospectively. Independent risk factors of 14-day mortality were determined by Cox regression analysis. A total of 99 patients with non-carbapenemase-producing CRKP infections were treated, and 14-day mortality was 27.3%. Among 67 patients treated with appropriate antimicrobial therapy, most ($n = 61$) patients received monotherapy. The 14-day mortality was lower in patients treated with appropriate monotherapy (21.3%) than in those treated with inappropriate therapy (37.5%). The multivariate regression model identified monotherapy (hazard ratio [HR], 0.30; 95% confidence interval [CI], 0.13–0.71; $P = 0.005$) as protective factor, and APACHE II scores (HR, 1.09; 95% CI, 1.01–1.18; $P = 0.02$) as factor associated with 14-day mortality. Tigecycline, colistin, and carbapenem were the most commonly used drugs. This study provides evidence supporting the efficacy of monotherapy in the treatment of non-carbapenemase-producing CRKP infections, and provides a future target for antibiotics stewardship for CRKP infection.

Chin-Fang Su and Chien Chuang contributed to this manuscript equally.

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99 non-CP-CRKP (2013.1-2014.12, 16 sites)

- ESBL: 88 SHV, 60 TEM, 45 CTX-M, 17 OXA-1.
- AmpC: 60 DHA-1, 5 CMY-2.
- 54 loss of OmpK35/36 porin, 36 OmpK35, 4 OmpK36

- 14-day mortality 27.3%
- 67.7% (67/99) appropriate antibiotic tx
- 61.6% (61/99) received monotherapy (Tige, Colistin, Carbapenem)
- Independent factors for mortality: monotherapy, appropriate antibiotics & APACHEII

Polymyxins

- Polymycin B, Polymycin E (colistin, prodrug-colistimethate sodium CMS)
- Mechanism: bind to negatively charged phosphate moieties in the lipid A fracture of LPS (GNB), disruption of cell membrane
- A very narrow therapeutic window: 2 µg/ml
- Nephrotoxic (2.5µg/ml) and neurotoxic
- 13.5-31.7% of colistin resistant CRE-KP (e.g. modification in lipid A, overexpression protein OprH, alterations in the mgrB gene...)

Landersdorfer CB *Semin Respir Crit Care Med* 2015;36:126. Pena I *Int J Antimicrob Agent* 2014;43:460

Tigecycline

- Mechanism: bacteriostatic, interferes with protein synthesis by binding to 30S ribosomal subunit
- A large volume of distribution (7-10L/kg), 40h of half-life, excreted into feces via bile, very little excreted in urine
- Dosage: 0.60ug/ml after a 100mg infusion, activity based on $AUC/MIC > 12.5$
- High dose : 100mg q12h for VAP
- CRE-KPC/MBLs: $MIC_{90} = 1\mu g/ml$, in combination with colistin and carbapenem
- Resistance: multidrug efflux pump (mutation in ramA....)

Castanheira M *Antimicrob Agents Chemother* 2008;52:570. De Pascale G *Crit Care* 2014;18:R90

Fosfomycin

- Mechanism: inhibits the first step in synthesis of peptidoglycan by blocking the formation of N-acetylmuramic acid through competitive inhibition of phosphoenol pyruvate synthetase.
- Antimicrobial spectrum: GPC, GNB
- >50% bioavailable after oral administration, largely excreted unchanged into the urine (e.g. UTI, prostatitis)
- 4g q6h with high-serum concentration
- 20% resistance rate (e.g. fosfomycin-inactivating enzyme..)
- Combination use only

Gobernade M. Rev Esp Quimioter 2003;16:15, Michalopoulos AS Int J Infect Dis 2011;15:e732. Pontikis K. Int J Antimicrob Agents 2014;43:52

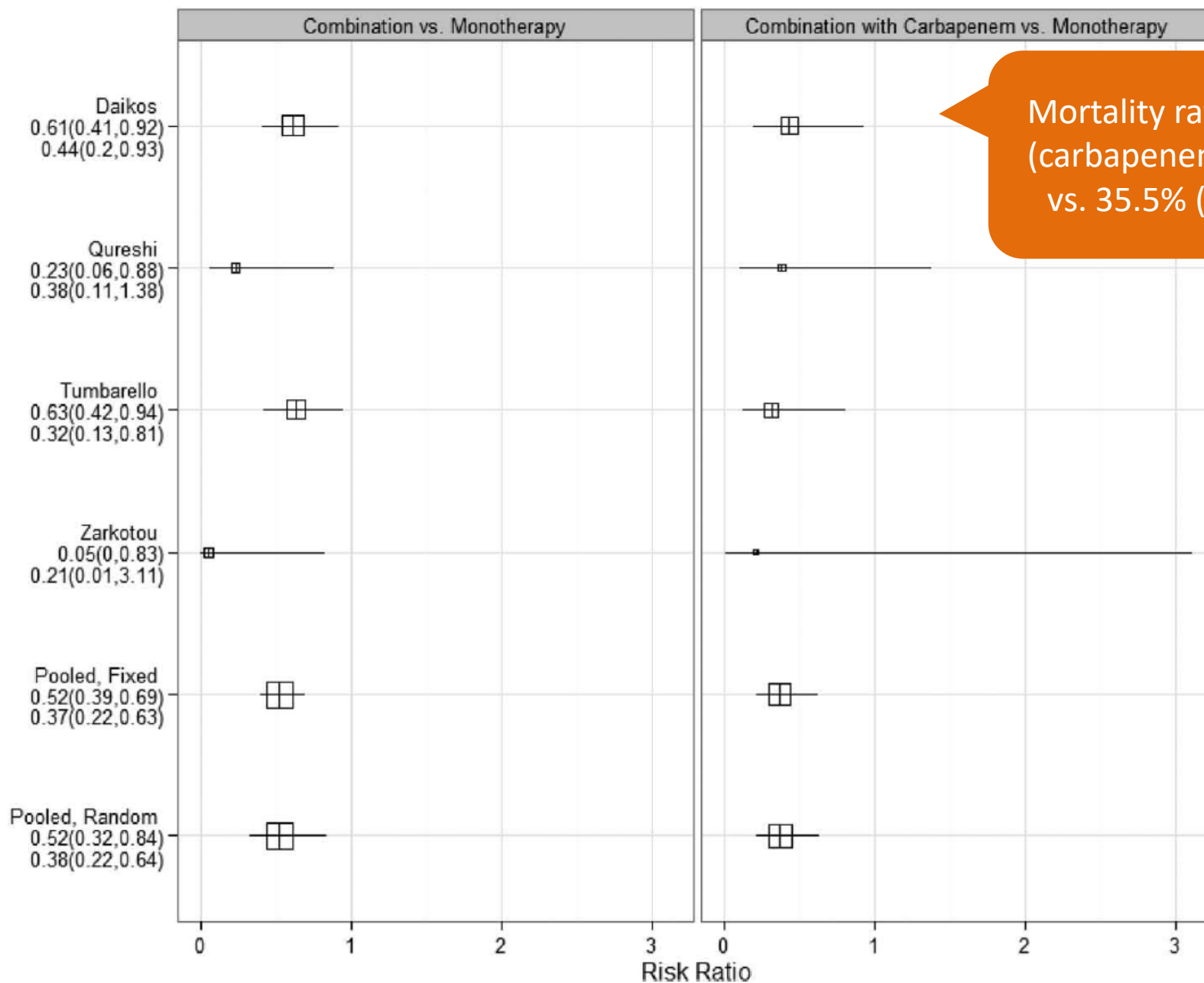
Aminoglycosides

- Mechanism: inhibition of protein synthesis (reduction in carbapenaemase production) + beta-lactam (cell wall synthesis inhibitor)
- Nephrotoxic and ototoxic

Combination or Monotherapy

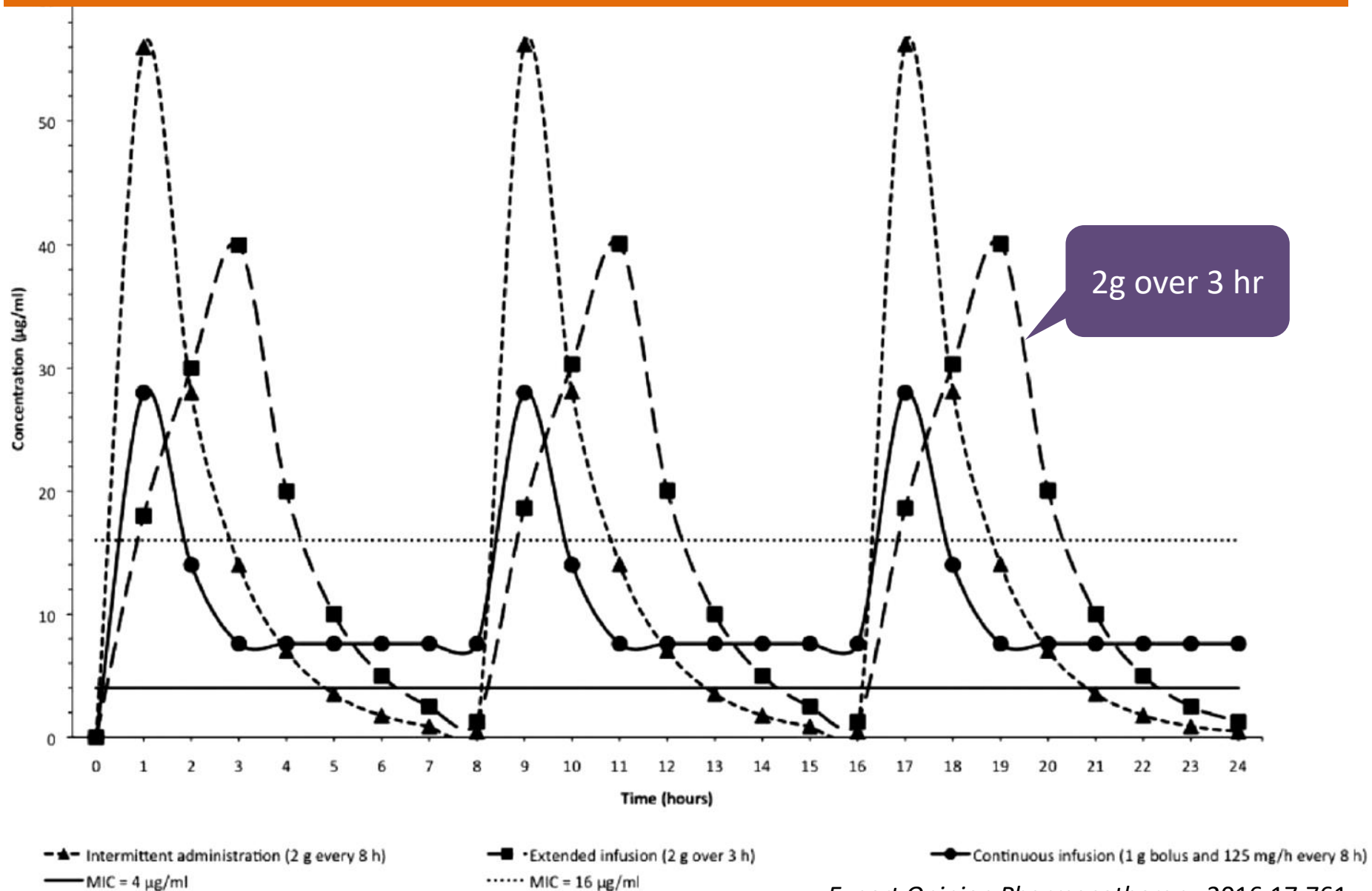
- **Carbapenem** with colistin (polymyxin B): synergy rate 55%-67%
- **Carbapenem** + fosfomycin: synergy 70%
- **Colistin** (polymyxin B) + rifampin, doxycycline, tigecycline
- **Ertapenem + doripenem**

Zusman O Antimicrob Agents Chemother 2013;57:5104. Jernigan MG Antimicrob Agents Chemother 2012;56:3395. Souli M Antimicrob Agents Chemother 2011;55:2395



Mortality rate: 19.3%
(carbapenem MIC≤8)
vs. 35.5% (MIC >8)

Concentration of meropenem: bactericidal activity by concentration above MIC for 40-50% of time



Organism	Resistant to	Primary treatment	Alternative treatment	Comments
ESBL Produced by Enterobacteriaceae, PA	All ceph, FQ, AG, TMP-SMX	MER or IMP	Ceftolozane- tazobactam Ceftazidime- avibactam Meropenem- vaborbactam	Pip-tazo discordance For UTI: Nitrofurantoin, Fosfomycin
OXA-48 producing Enterobacteriaceae	All PCNs, FQ, AG, TMP-SMX	If sensitive: Cefepime, Ceftazidime	If critically: Ceftazidime- avibactam, Aztreonam	Mediterranean sea
KPC producing Enterobacteriaceae	All PCNs, cepha, aztreonam, carbapne ms, FQ, AG, TMP-SMX	Ceftaz-avi, meropenem- vaborbactam	MER + Polymycin B	For pneumonia: add nebulized colistin 50-75mg bid ERTA + MER (JAC 69:1718,2015)
Metallo- carbapenemase		Ceftaz-avi + aztreonam		Table 5B: Treatment options for MDR (熱病2019)

- Lin YT. *Open Forum Infectious Dis*

Risk Factors for Treatment Failure of Polymyxin B Mono- Carbapenem-Resistant *Klebsiella pneumoniae* Infections

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Polymyxins are reserved for salvage therapy of infections caused by carbapenem-resistant *Klebsiella pneumoniae* (CRKP). Though synergy has been demonstrated for the combination of polymyxins with carbapenems or tigecycline, these tests are nonstandardized, and the clinical effect of synergy remains unclear. This study describes outcome of CRKP infections who were treated with polymyxin B monotherapy. We retrospectively reviewed the medical records of patients with CRKP infections who received polymyxin B monotherapy from 2007 to 2011. Clinical, microbiology, and treatment data were collected. Risk factors for treatment failure were identified by logistic regression. Forty patients were included in the analysis. Twenty-nine of 40 (73%) patients achieved clinical cure as defined by clinician documentation of resolution of signs and symptoms of infections, and 17/32 (53%) patients with follow-up culture data achieved microbiologic cure. Of-treatment mortality was 10%, and 30-day mortality was 28%. In a multivariate analysis, baseline renal insufficiency was associated with a 6.0-fold increase in clinical failure after adjusting for septic shock (odds ratio [OR] = 6.0; 95% confidence interval [CI] = 1.22 to 29.59). Breakthrough infections with organisms intrinsically resistant to polymyxins occurred during the treatment. Eighteen of 40 (45%) patients developed a new CRKP infection a median of 23 days after initial polymyxin B treatment, and 3 of these 18 infections were polymyxin resistant. The clinical cure rate achieved in this retrospective study was 73% of patients with CRKP infections treated with polymyxin B monotherapy. Baseline renal insufficiency was a risk factor for treatment failure after adjusting for septic shock. Breakthrough infections with organisms with intrinsic resistance to polymyxins and development of resistance to polymyxin B in subsequent CRKP isolates are of concern.

Infections caused by carbapenem-resistant *Klebsiella pneumoniae* (CRKP) have been associated with poor outcomes. An overall mortality rate of 48% is reported for patients with CRKP, compared to 26% for those with carbapenem-susceptible *K. pneumoniae* infections (1). CRKP isolates confer resistance through the production of *K. pneumoniae* carbapenemases (KPCs), which are able to hydrolyze all currently available β -lactams, including carbapenems (2). Due to additional mechanisms of resistance commonly found in KPC-producing isolates, they often demonstrate resistance to other antimicrobial classes, including fluoroquinolones and aminoglycosides (2, 3). This limits therapeutic choices, as polymyxins (polymyxin B or colistin) and tigecycline may be the only antibiotics that retain *in vitro* and *in vivo* microbiological activity.

CRKP infections are often treated with combination therapy (4, 5); however, the choice of agents is both limited and challenging. Combination therapy with aminoglycosides, if susceptibility is preserved, carries an increased risk of nephrotoxicity. Due to a high volume of distribution, low blood levels, and high MIC, tigecycline is a "last resort" option, particularly for bloodstream infections. High MICs of carbapenems preclude achieving therapeutic levels even with pharmacodynamically optimized dosing strategies. Although *in vitro* synergy against CRKP has been observed when polymyxin B is used in combination with rifampin, cefepime, aminoglycosides, carbapenems, or tigecycline, testing methods are nonstandardized, and therefore the clinical effect of synergy remains unclear (3, 6–8). Limited data have demonstrated survival benefits of combination therapy using polymyxin B or colistin with carbapenems and/or tigecycline for patients with CRKP bacteremia (4, 5). Given the frequently encountered challenges of combination therapy, we aimed to describe the out-

comes for patients with CRKP infections treated with polymyxin B monotherapy.

MATERIALS AND METHODS

Study design. We conducted a retrospective review of medical records of patients with CRKP infections treated with polymyxin B monotherapy from 2007 to 2011. Patients were identified from the New York University Hospital database. Patients with CRKP infections who received polymyxin B monotherapy for at least 48 h were included in the study.

Definitions. Clinical cure was defined as resolution of signs and symptoms of infection, and microbiologic cure was defined as a negative culture result.

Data collection. Clinical, microbiology, and treatment data were collected from medical records.

Statistical analysis. Data were analyzed using STATA 11.0 (College Station, TX). Descriptive statistics were calculated for all variables. Logistic regression was used to identify risk factors for treatment failure.

Ethics approval. The study was approved by the Institutional Review Board at New York University.

40 CR-KP (50 KP, 14 E.coli, 16 sites, **2007-2012**)

- Carbapenemases and non-CPKP: ?
- Median age 76 yr, 14 bacteremia, 12 UTI, 7 pneumonia, 4 STSI, 2 IAI, 1 osteomyelitis
- Receiving polymyxin B monotherapy

- Clinical cure: 73% (29/40)
- 30-day mortality = 28%
- Independent treatment failure: baseline renal insufficiency AOR 6.0 (1.22-29.59) &
- Polymyxin resistance during tx = 7.5% (3/40)
- **New CRKP infection in 45% (18/40) at a median 23 days after initial polymyxin B tx, 37.5% (3/18) with polymyxin B resistance**

Conclusions

- Mortality rate of Bloodstream infections of CRE : 50%
- Current available antibiotics for CRE: colistin, tigecycline, fosfomycin, AK
- A combination therapy for CRE (e.g. carbapenem MIC < 16 ug/ml and prolonged infusion + colistin / tige)
- CRE genotyping (A, B, D ?)
- Molecular surveillance study and Amber classification for individualized treatment of CRE